

Carbohydrate Research 271 (1995) 247-251

# Note

# Synthesis of benzyl O-(3-O-methyl- $\beta$ -D-galactopyranosyl)-(1 $\rightarrow$ 3)-2-acetamido-2-deoxy- $\alpha$ -D-galactopyranoside and benzyl O-( $\beta$ -D-galactopyranosyl)-(1 $\rightarrow$ 3)-2-acetamido-2-deoxy-6-O-methyl- $\alpha$ -D-galactopyranoside as specific acceptors for sialyltransferases $^{\frac{1}{\alpha}}$

Rakesh K. Jain, Conrad F. Piskorz, E.V. Chandrasekaran, Khushi L. Matta \*

Department of Gynecologic Oncology, Roswell Park Cancer Institute, Elm & Carlton Streets, Buffalo, NY 14263, USA

Received 7 September 1994; accepted in revised form 14 January 1995

Keywords: Sialyltransferases

In our laboratory, we are currently involved in the synthesis of specific acceptors for sialyltransferases (STs). GalNAc:  $\alpha$ -(2  $\rightarrow$  6) STs catalyze the transfer of a sialic acid to 0-6 of GalNAc, and Gal:  $\alpha$ -(2  $\rightarrow$  3) STs are involved in the synthesis of oligosaccharides containing sialic acid linked  $\alpha$ -(2  $\rightarrow$  3) to Gal residues. Based on the specificities of different acceptor-substrates and differences in biochemical properties between enzymes from different sources, at least 12 different sialyltransferases (STs) are known [2–5].

In order to achieve a specific, quantitative determination of individual STs, we embarked on a synthetic program to obtain well defined, low molecular weight oligosaccharides capable of acting as acceptors for a single enzyme, even in the presence of other related enzymes. The present communication demonstrates that 2,4,6-tri-O-acetyl-3-O-methyl- $\alpha$ -D-galactopyranosyl bromide (2) is an effective glycosylating reagent for the synthesis of benzyl O-(3-O-methyl- $\beta$ -D-galactopyranosyl)-(1  $\rightarrow$  3)-2-acetamido-2-deoxy- $\alpha$ -D-galactopyranoside (7) and benzyl O-(3-O-methyl- $\beta$ -D-galactopyranosyl)-

<sup>\*</sup> Synthetic Studies in Carbohydrates, Part 96. For Part 95, see ref. [1].

<sup>\*</sup> Corresponding author.

 $(1 \rightarrow 3)$ -2-acetamido-2-deoxy-6-O-methyl- $\alpha$ -D-galactopyranoside (9). We have also accomplished the synthesis of benzyl  $O(\beta$ -D-galactopyranosyl)- $(1 \rightarrow 3)$ -2-acetamido-2-deoxy-6-O-methyl- $\alpha$ -D-galactopyranoside (5). Compounds 5 and 7 should serve as ideal acceptors in measuring the O-glycan related  $\alpha$ - $(2 \rightarrow 3)$  and  $\alpha$ - $(2 \rightarrow 6)$  STs in tumors, other malignant tissues, as well as in cultured human cells for the purpose of studying biosynthetic pathways. Compound 9 may serve as an inhibitor in the study of this class of enzymes.

Methylation of benzyl O-(2,3,4,6-tetra-O-acetyl- $\beta$ -D-galactopyranosyl)-(1  $\rightarrow$  3)-2-acetamido-2-deoxy- $\alpha$ -D-galactopyranoside (3) [6] with trimethyloxonium tetrafluoroborate-2,6-di-(*tert*-butyl)-4-methyl pyridine [7] in dichloromethane gave the 6-O-methyl derivative 4 in 82% yield. Zemplen transesterification of 4 furnished compound 5 in 75% yield. The <sup>13</sup>C NMR spectrum of amorphous 5 was in agreement with the structure assigned (see Experimental section).

A common intermediate, namely, benzyl O-(2,4,6-tri-O-acetyl-3-O-methyl- $\beta$ -D-galactopyranosyl)-(1  $\rightarrow$  3)-2-acetamido-2-deoxy- $\alpha$ -D-galactopyranoside (6) was used for the synthesis of 7 and 9.

2,4,6-Tri-O-acetyl-3-O-methyl- $\alpha$ -D-galactopyranosyl bromide 2, obtained through acetolysis of methyl 2,4,6-tri-O-acetyl-3-O-methyl- $\alpha$ -D-galactopyranoside [8] (1), followed by treatment with 31% HBr in glacial acetic acid, was the key glycosyl donor. Regioselective methylation [9,10] of methyl  $\alpha$ -D-galactopyranoside followed by acetoly-

sis with 1% H<sub>2</sub>SO<sub>4</sub> in acetic anhydride provided compound 1 in 72% yield. Glycosylation of benzyl 2-acetamido-4,6-O-benzylidene-2-deoxy-α-D-galactopyranoside with bromide 2 in 1:1 benzene-nitromethane and in the presence of Hg(CN)2, followed by removal of the acetal ring with 70% aqueous acetic acid, produced the diol 6 in 77% yield, O-Deacetylation of 6 with methanolic sodium methoxide afforded, in 90% yield, the desired disaccharide 7. A similar methylation of 6 with trimethyloxonium tetrafluoroborate-2,6-di-(tert-butyl)-4-methyl pyridine and processing in a manner analogous to that described for 3 (to give 5) afforded compound 9 in 51% yield. The <sup>13</sup>C NMR spectra and FABS (see Experimental section) of both 7 and 9 were in agreement with the structures assigned.

In the  $^{13}$ C NMR spectra of compounds 5 and 9, the C-6 resonance of the GalNAc  $\alpha$ -OBn residue exhibited a downfield shift ( $\delta$  67.76 and 67.73), thus confirming this position as the site of methylation. Similarly, the resonance of C-3 of the Gal residue in compounds 7 and 9 displayed a downfield shift at  $\delta 83.03$  confirming this position as the site of methylation in these compounds.

# 1. Experimental

General methods. — Optical rotations were measured at ~ 25°C with a Perkin-Elmer 241 Polarimeter. TLC was conducted on glass plates, precoated with a 0.25 mm layer of Silica Gel 60F-254 (Analtech GHLF uniplates). The compounds were visualized by exposure to UV light and/or by spraying with 5% H<sub>2</sub>SO<sub>4</sub> in EtOH and charring. The silica gel used for column chromatography was Baker Analyzed (60-200 mesh). NMR spectra were recorded at ~25°C; <sup>1</sup>H NMR spectra were obtained with a Varian EM-390 (90 MHz) and with a Bruker AM-400 (400 MHz), and <sup>13</sup>C NMR spectra with a Bruker AM-400 spectrometer operating at 100.6 MHz for <sup>13</sup>C. All chemical shifts are referenced to tetramethylsilane (TMS). Solutions in organic solvents were generally dried with anhyd Na<sub>2</sub>SO<sub>4</sub>. Dichloromethane, DMF and benzene were kept dry over 4 Å molecular sieves. Elemental analyses were performed by the Robertson Laboratory, Madison, New Jersey.

1,2,4,6-Tetra-O-acetyl-3-O-methyl-D-galactopyranose (1). — A solution of methyl 3-O-methyl-α-D-galactopyranoside (10 g) in acetic anhydride (150 mL) containing ~ 1% by volume of concd H<sub>2</sub>SO<sub>4</sub> was stirred for 6 h at room temperature. The mixture was then diluted with dichloromethane (700 mL) and successively washed with water, saturated NaHCO<sub>3</sub> and water, dried and concentrated under reduced pressure. The residue was applied to a column of silica gel and eluted with 30% ethyl acetate in hexane to give 1 (11.5 g, 71.6%);  $[\alpha]_D + 109 (c 1.7, CHCl_3)$ ; <sup>1</sup>H NMR (CDCl<sub>3</sub>):  $\delta 6.29$ (d, 0.9 H,  $J \sim 4$  Hz), 3.40 (bs, 3 H, OMe), 2.17–2.03 (cluster of s, 12 H,  $4 \times$  OAc).

Anal. Calcd for C<sub>14</sub>H<sub>22</sub>O<sub>9</sub>: C, 50.29; H, 6.63. Found: C, 50.15; H, 6.59.

2,4,6-Tri-O-acetyl-3-O-methyl- $\alpha$ -D-galactopyranosyl bromide (2). — 1,2,4,6-Tetra-O-acetyl-3-O-methyl-p-galactopyranose (30 g) in methylene chloride (300 mL) and acetic anhydride (15 mL) was cooled (0°C, bath) and treated with 31% HBr in glacial acetic acid (200 mL), and the mixture stirred for 2 h at 0°C. The mixture was diluted with methylene chloride (200 mL), and successively washed with cold water, cold saturated NaHCO<sub>3</sub> and cold water, dried and evaporated. The residue was dissolved in methylene chloride, and addition of ether-hexane resulted in the precipitation of 2 (21 g, 59%), a white powder;  $[\alpha]_D + 202 (c \ 0.5, CHCl_3)$ ; <sup>1</sup>H NMR (CDCl<sub>3</sub>):  $\delta 5.62 (d, J = 4)$  Hz, 1 H, H-1), 5.52 (d, J = 3 Hz, 1 H, H-4), 4.87 (dd, J = 4.5 Hz, 1 H, H-3), 3.43 (s, 3 H, OMe), 2.13, 2.10 and 2.07 (each s, 9 H,  $3 \times OAc$ ).

Benzyl O-(2,3,4,6-tetra-O-acetyl- $\beta$ -D-galactopyranosyl)- $(1 \rightarrow 3)$ -2-acetamido-2deoxy-6-O-methyl- $\alpha$ -D-galactopyranoside (4). — To a cold (0°, bath) and stirred solution of benzyl O-(2,3,4,6-tetra-O-acetyl- $\beta$ -D-galactopyranosyl)-(1  $\rightarrow$  3)-2acetamido-2-deoxy-α-p-galactopyranoside (1.2 g, 1.9 mmol) in methylene chloride (30 mL) was added 2,6-di-(tert-butyl)-4-methylpyridine (0.8 g, 3.9 mmol) and trimethyloxonium tetrafluoroborate (0.4 g, 2.7 mmol). Stirring was continued for 5 h at the same temperature. The mixture was diluted with methylene chloride and successively washed with saturated aqueous NaHCO<sub>3</sub>, water, dried and concentrated under reduced pressure. The residue was applied to a column of silica gel and eluted with a solvent gradient consisting of 3-5% methanol in chloroform. The earlier fractions contained the fastermigrating compound 4. On concentration these fractions afforded a solid (0.75 g, 82% on the basis of 3 reacted);  $[\alpha]_D + 85^\circ$  (c 0.6, CHCl<sub>3</sub>); <sup>1</sup>H NMR (CDCl<sub>3</sub>):  $\delta 7.36 - 7.26$ (m, 5 H, aromatic), 5.46 (d, J = 9.6 Hz, 1 H, NH), 5.36 (d, J = 3.4 Hz, 1 H, H-4'), 5.18 (dd, J = 7.9 Hz, 1 H, H-2'), 4.98 (d, J = 3.5 Hz, 1 H, H-1), 4.94 (dd, 1 H, H-3'), 4.59 (d, J = 7.8 Hz, 1 H, H-1'), 3.41 (s, 3 H, OMe), 2.15, 2.05, 2.03, 1.97 (each s, 12 H,  $4 \times OAc$ ), 1.92 (s, 3 H, NAc); <sup>13</sup>C NMR (CDCl<sub>3</sub>):  $\delta$ 101.59 (C-1'), 97.42 (C-1), 78.09 (C-3), 68.43 (C-6), 61.43 (C-6'), 59.33 (OMe), 47.84 (C-2).

Anal. Calcd for  $C_{30}H_{41}NO_{15}$ : C, 54.95; H, 6.30; N, 2.14. Found: C, 54.82; H, 6.43; N, 1.94.

Later fractions contained pure unreacted compound 3 (0.3 g).

Benzyl O-(β-D-galactopyranosyl)-(1  $\rightarrow$  3)-2-acetamido-2-deoxy-6-O-methyl-α-D-galactopyranoside (5). — Compound 4 (0.7 g) was stirred in 0.05 M methanolic sodium methoxide (50 mL) for 16 h at room temperature. The solution was deionized with Amberlite IR-120 (H<sup>+</sup>) cation-exchange resin, filtered and concentrated under reduced pressure. After purification over a silica gel column with CHCl<sub>3</sub>-MeOH-H<sub>2</sub>O (13:6:1) as the eluent, 5 (0.39 g, 75%) was obtained as an amorphous solid, [α]<sub>D</sub> +125° (c 1.2, DMSO); <sup>1</sup>H NMR [(CD<sub>3</sub>)<sub>2</sub>SO]: δ7.63 (d, J = 8.1 Hz, 1 H, NH), 7.36-7.35 (m, 5 H, aromatic), 4.80 (d, J = 3.5 Hz, 1 H, H-1), 3.27 (s, 3 H, OMe), 1.82 (s, 3 H, NAc); <sup>13</sup>C NMR: δ103.76 (C-1'), 96.63 (C-1), 75.64 (C-3), 75.38 (C-5'), 73.37 (C-5), 71.99 (C-3'), 69.37 (C-2'), 68.35 (C-4), 68.23 (C-4'), 67.76 (C-6), 60.60 (C-6'), 58.40 (OMe), 48.45 (C-2), 22.76 (NAc); m/z: 488.2 (M + H)<sup>+</sup>, 486.5 (M - H)<sup>-</sup>.

Anal. Calcd for  $C_{22}H_{33}NO_{11}$ : C, 54.20; H, 6.82; N, 2.87. Found: C, 53.94; H, 7.11; N, 2.84.

Benzyl O-(2,4,6-tri-O-acetyl-3-O-methyl- $\beta$ -D-galactopyranosyl)- $(1 \rightarrow 3)$ -2-acetamido-2-deoxy- $\alpha$ -D-galactopyranoside (6). — A stirred mixture of benzyl 2-acetamido-4,6-O-benzylidene-2-deoxy- $\alpha$ -D-galactopyranoside (1.6 g, 4 mmol) and powdered Hg(CN)<sub>2</sub> (2.0 g, 8 mmol) in 1:1 benzene—nitromethane (100 mL) was boiled until 20 mL of the solvent had distilled off. The temperature was then adjusted to 45°C, and a solution of 2,4,6-tri-O-acetyl-3-O-methyl- $\alpha$ -D-galactopyranosyl bromide (3.0 g, 7.8 mmol) in benzene (30 ml) was added and stirring was continued for 16 h at room temperature. The mixture was diluted with benzene and washed with saturated aqueous NaHCO<sub>3</sub>, 10% aqueous KI solution and water, dried and evaporated to a syrup. This crude compound in 70% aqueous acetic acid (100 mL) was stirred for 2 h at ~90°C. Acetic acid was evaporated under reduced pressure, the last traces being removed by co-evaporation with several added portions of toluene to leave a residue that was purified on a column of silica gel using a solvent gradient consisting of 3–5% methanol

in chloroform, yielding after evaporation of fractions amorphous product **6** (1.9 g, 77%);  $[\alpha]_D + 103^\circ$  (c 1.2, CHCl<sub>3</sub>); <sup>1</sup>H NMR (CDCl<sub>3</sub>):  $\delta$ 7.73–7.31 (m, 5 H, aromatic), 5.59 (d, J = 9.3 Hz, 1 H, NH), 5.45 (d, J = 3.1 Hz, 1 H, H-4'), 4.99 (d, J = 3.5 Hz, 1 H, H-1), 4.55 (d, J = 7.9 Hz, 1 H, H-1'), 3.34 (s, 3 H, OMe), 2.13, 2.08 and 2.04 (each s, 9 H, 3 × OAc), 1.94 (s, 3 H, NAc); <sup>13</sup>C NMR (CDCl<sub>3</sub>):  $\delta$ 101.46 (C-1'), 97.48 (C-1), 79.37 (C-3'), 77.64 (C-3), 62.72 (C-6'), 61.97 (C-6), 57.98 (OMe), 47.89 (C-2).

Anal. Calcd for  $C_{28}H_{39}NO_{14}$ : C, 54.80; H, 6.41; N, 2.28. Found: C, 55.11; H, 6.29; N, 2.14.

Benzyl O-(3-O-methyl-β-D-galactopyranosyl)-(1  $\rightarrow$  3)-2-acetamido-2-deoxy-α-D-galactopyranoside (7). — O-Deacetylation of compound 6 (0.5 g) with 0.025 M methanolic sodium methoxide afforded compound 7 (0.36 g, 90%);  $[\alpha]_D$  + 127° (c 1.0, Me<sub>2</sub>SO); <sup>1</sup>H NMR [(CD<sub>3</sub>)<sub>2</sub>SO]; δ7.60 (d, J = 8.0 Hz, 1 H, NH), 7.37–7.34 (m, 5 H, aromatic), 4.79 (d, J = 3.6 Hz, 1 H, H-1), 3.32 (s, 3 H, OMe), 1.82 (s, 3 H, NAc); <sup>13</sup>C NMR: δ103.64 (C-1'), 96.40 (C-1), 83.03 (C-3'), 75.83 (C-3), 75.12 (C-5'), 71.45 (C-5), 69.62 (C-2'), 68.02 (C-4), 67.36 (C-4'), 60.68 (C-6), 60.40 (C-6'), 56.15 (OMe), 48.48 (C-2), 22.63 (NAc); m/z: 488.3 (M + H)<sup>+</sup>, 486.4 (M – H)<sup>-</sup>.

Anal. Calcd for  $C_{22}H_{33}NO_{11}$ : C, 54.20; H, 6.82; N, 2.87. Found: C, 54.11; H, 6.91; N, 2.91.

Benzyl O-(3-O-methyl-β-D-galactopyranosyl)-(1  $\rightarrow$  3)-2-acetamido-2-deoxy-6-O-methyl-α-D-galactopyranoside (9). — Methylation of compound 6 (1.2 g), followed by O-deacetylation as described for the preparation of 5, afforded compound 9 (0.5 g, 51%) after silica gel column chromatography (solvent gradient consisting of 15–20% methanol in chloroform); [α]<sub>D</sub> +121° (c 1.6, Me<sub>2</sub>SO); <sup>1</sup>H NMR [(CD<sub>3</sub>)<sub>2</sub>SO]: δ7.61 (d, J = 7.9 Hz, 1 H, NH), 7.38–7.35 (m, 5 H, aromatic), 4.80 (d, J = 3.5 Hz, 1 H, H-1), 3.32 and 3.27 (each s, 6 H, 2 × OMe), 1.81 (s, 3 H, NAc); <sup>13</sup>C NMR: δ103.64 (C-1'), 96.58 (C-1), 83.03 (C-3'), 75.54 (C-3), 75.16 (C-5'), 71.93 (C-5), 69.60 (C-2), 69.33 (C-4), 68.27 (C-4'), 67.73 (C-6), 60.45 (C-6'), 58.34 and 56.17 (2 × OMe), 48.37 (C-2), 22.63 (NAc); m/z: 502.3 (M + H)<sup>+</sup>, 500.5 (M – H)<sup>-</sup>.

Anal. Calcd for  $C_{23}H_{35}NO_{11}$ : C, 55.08;H, 7.03; N, 2.79. Found: C, 54.94; H, 7.10; N, 2.64.

## Acknowledgements

This investigation was supported by Grant No. CA35329 awarded by the National Cancer Institute.

### References

- [1] R.K. Jain, X-G. Liu and K.L. Matta, Carbohydr. Res., 268 (1995) 279-285.
- [2] E. Roos, Biochim. Biophys. Acta, 738 (1984) 263-284.
- [3] J.C. Paulson and K.J. Colley, J. Biol. Chem., 264 (1989) 17615-17618.
- [4] J. Weinstein, E.W. Lee, K. McEntee, P.H. Lai and J.C. Paulson, J. Biol. Chem., 262 (1987) 17735-17743.
- [5] W. Gillespie, S. Kelm, and J.C. Paulson, J. Biol. Chem., 267 (1992) 21004-21010.
- [6] C.F. Piskorz, S.A. Abbas and K.L. Matta, Carbohydr. Res., 126 (1984) 115-124.
- [7] P. Fugedi and J. Kovacs, Int. Carbohydr. Symp., XVth (1990) 77.
- [8] H.M. Flowers, Carbohydr. Res., 39 (1975) 245-251.
- [9] A.S. Shaskov, A.I. Vsov, S.V. Yarotskii and A.B. Rabovskii, *Bioorg. Khim.*, 4 (1978) 1489–1494; *Sov. J. Bioorg. Chem.* (Engl. Transl.), 4 (1978) 1071–1076.
- [10] W. Liao, Y. Liu and D. Lu, Carbohydr. Res., 260 (1994) 151-154.